

Chapter 2. Development and disease.

Our understanding of developmental mechanisms has been transformed during the 20th century by advances in genetic technology and molecular biology. The key genes regulating embryonic segmentation and the allocation of regional fate were identified in *Drosophila*^{1 2 3 4 5 6 7} and are highly conserved in other segmented organisms. An additional set of genes regulating epithelial planar cell polarity (PCP) has been studied^{8 9 10}, although mainly during later development. The relationship between embryonic patterning and cellular polarity remains unclear. Recently, novel roles for the core PCP genes have been identified in neuronal function and disease mechanisms, both in model organisms and humans. For example, the mammalian Prickle (Pk) orthologues (Pk-1, -2 and -3) are expressed during embryonic convergent extension movements, somite formation and left-right symmetry breaking; with later functions during limb growth^{11 12 13 14}. In the zebrafish, Pk orthologues are required for cell motility during gastrulation and neuronal migration¹⁵. Pk-associated disease syndromes include autism spectrum disorders, myoclonus epilepsy, lissencephaly and cancer metastasis^{16 17 18 19 20 21 22 23 24}; similar defects are associated with other PCP mutants, including neural tube malformations in *Celsr1* and *Celsr3*^{25 26 27}. Similarly, Testin (Tes) has a female germline function and acts as a tumour suppressor in mice and humans^{28 29}. Thus, PCP gene functions regulate complex developmental processes and are associated with multiple disease syndromes. However, the PCP genes do not form a discrete group, and their associated mutant phenotypes depend on interactions between transcription factors (TFs), growth factors (GFs) and morphogens. Regulation of none of these genetic functions is independent of the others.

Summary:

Multiple morphogenetic defects and adult disease syndromes are associated with mis-regulation of PCP signalling interactions. These processes are co-ordinated between individual cells, across epithelial fields, and maintained within adult tissues.

References:

1. Lewis, E. B. A gene complex controlling segmentation in *Drosophila*. *Nature* 276, 565 (1978).
2. Nusslein-Volhard, C. & Wieschaus, E. Mutations affecting segment number and polarity in *Drosophila*. *Nature* 287, 795–801 (1980).
3. Nusslein-Volhard, C., Wieschaus, E. & Kluding, H. Mutations affecting the pattern of the larval cuticle in *Drosophila melanogaster*. *Roux's Arch. Dev. Biol.* 193, 267–282 (1984).
4. Wieschaus, E., Nusslein-Volhard, C. & Jurgens, G. Mutations affecting the pattern of the larval cuticle in *Drosophila melanogaster*. *Roux's Arch. Dev. Biol.* 193, 296–307 (1984).
5. Anderson, K. V., Jurgens, G. & Nusslein-Volhard, C. Establishment of dorsal-ventral polarity in the *Drosophila* embryo. Genetic studies on the role of the Toll gene product. *Cell* 42, 779–789 (1985).
6. Lewis, E. B. Regulation of the genes of the Bithorax complex in *Drosophila*. *Cold Spring Harb. Symp. Quant. Biol.* 50, 155–164 (1985).
7. Levine, M., Hafen, E., Garber, R. L. & Gehring, W. J. Spatial distribution of Antennapedia transcripts during *Drosophila* development. *EMBO J.* 2, 2037–2046 (1983).

8. Gubb, D. & Garcia-Bellido, A. A genetic analysis of the determination of cuticular polarity during development in *Drosophila melanogaster*. *J. Embryol. Exp. Morphol.* 68, 37–57 (1982).
9. Vinson, C. R. & Adler, P. N. Directional non-cell autonomy and the transmission of polarity information by the frizzled gene of *Drosophila*. *Nature* 329, 549–551 (1987).
10. Strutt, D. & Strutt, H. Differential activities of the core planar polarity proteins during *Drosophila* wing patterning. *Dev. Biol.* 302, 181–194 (2007).
11. Torban, E., Kor, C. & Gros, P. Van Gogh-like2 (Strabismus) and its role in planar cell polarity and convergent extension in vertebrates. *Trends Genet.* 20, 570–577 (2004).
12. Cooper, O., Sweetman, D., Wagstaff, L. & Munsterberg, A. Expression of avian prickle genes during early development and organogenesis. *Dev Dyn* 237, 1442–8 (2008).
13. Bekman, E. & Henrique, D. Embryonic expression of three mouse genes with homology to the *Drosophila melanogaster* prickle gene. *Mech Dev* 119 Suppl 1, S77–81 (2002).
14. Minegishi, K. *et al.* A Wnt5 Activity Asymmetry and Intercellular Signaling via PCP Proteins Polarize Node Cells for Left-Right Symmetry Breaking. *Dev. Cell* 40, 439–452.e4 (2017).
15. Carreira-Barbosa, F. *et al.* Prickle 1 regulates cell movements during gastrulation and neuronal migration in zebrafish. *Development* 130, 4037–46 (2003).
16. Manak, J. R. *et al.* Mutations in Prickle orthologs cause seizures in flies, mice, and humans. *Am. J. Hum. Genet.* 88, 138–149 (2011).
17. Veeman, M. T., Slusarski, D. C., Kaykas, A., Louie, S. H. & Moon, R. T. Zebrafish prickle, a modulator of noncanonical Wnt/Fz signaling, regulates gastrulation movements. *Curr Biol* 13, 680–5 (2003).
18. Bassuk, A. G. *et al.* A Homozygous Mutation in Human PRICKLE1 Causes an Autosomal-Recessive Progressive Myoclonus Epilepsy-Ataxia Syndrome. *Am. J. Hum. Genet.* 83, 572–581 (2008).
19. Ehaideb, S. N. *et al.* prickle modulates microtubule polarity and axonal transport to ameliorate seizures in flies. *Proc Natl Acad Sci U S A* 111, 11187–92 (2014).
20. Daulat, A. M. *et al.* Prickle1 contributes to cancer cell dissemination through its interaction with mTORC2. *Dev. Cell* 37, 311–325 (2016).
21. Bassuk, A. G. & Sherr, E. H. A de novo mutation in Prickle1 in fetal agenesis of the corpus callosum and polymicrogyria. *J. Neurogenet.* 29, 174–177 (2015).
22. Asad, M. *et al.* FZD7 drives in vitro aggressiveness in Stem-A subtype of ovarian cancer via regulation of non-canonical Wnt/PCP pathway. *Cell Death Dis.* 5, e1346 (2014).
23. Lim, B. C. *et al.* Prickle1 promotes focal adhesion disassembly in cooperation with the CLASP–LL5 β complex in migrating cells. *J. Cell Sci.* 129, 3115 (2016).
24. Kacey VanderVorst *et al.* Wnt/PCP signaling mediates breast cancer metastasis by promoting pro-invasive protrusion formation in collectively motile leader cells. *bioRxiv* 2022.01.07.475316 (2022)
25. Formstone, C. J., Moxon, C., Murdoch, J., Little, P. & Mason, I. Basal enrichment within neuroepithelia suggests novel function(s) for Celsr1 protein. *Mol. Cell. Neurosci.* 44, 210–222 (2010).
26. Murdoch, J. N. *et al.* Genetic interactions between planar cell polarity genes cause diverse neural tube defects in mice. *Dis. Model. Mech.* 7, 1153–1163 (2014).
27. Oozer, F., Yates, L. L., Dean, C. & Formstone, C. J. A role for core planar polarity proteins in cell contact-mediated orientation of planar cell division across the mammalian embryonic skin. *Sci. Rep.* 7, 1880–1880 (2017).

28. Drusco, A. *et al.* Knockout mice reveal a tumor suppressor function for Testin. *Proc. natl. Acad. Sci. U. S. A.* 102, 10947–10951 (2005).
29. Weeks, R. J., Kees, U. R., Song, S. & Morison, I. M. Silencing of TESTIN by dense biallelic promoter methylation is the most common molecular event in childhood acute lymphoblastic leukaemia. *Mol. Cancer* 9, 163–163 (2010).